Antibacterial Studies of Essential Oil and Different Extract Fractions from the Seeds of Nigella sativa L.

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ABSTRACT

The antibacterial activity of the neutral, acidic, basic fractions and essential oil of Nigella sativa seeds were tested for their in vitro activity against eight species of bacteria, Staphylococcus aureus, Bacillus subtilis, Proteus mirabilis, Salmonella typhi, E. coli, Pseudomonas aeruginosa, Salmonella typhimurium and Shigella flexneri were investigated and compared with standard antibiotics Gentamycin and Amoxicillin.

The results indicate that these essential oil, acidic and basic fractions have antibacterial activity against most of the tested bacteria. We
conclude from the above results, that the essential oil, acidic and basic fractions seeds of Nigella sativa have antibacterial activity against gram negative bacterial especially (Sal. typhimurium, Sal. typhi and Sh. flexneri) more than other gram negative and gram positive bacteria. The acidic and basic fractions compounds were confirmed spectrochemically by using the IR spectroscopy.

INTRODUCTION

Nigella sativa (black cumin) is an annual herbaceous plant growing in Western Asia and the Mediterranean region for its seeds. The seeds contain 40% volatile oil (1). The seeds of Nigella sativa have been used traditionally for centuries in the Middle East, Northern Africa and South Asia for the treatment of various diseases (2,3).

The plant extracts and essential oil showed a broad range of pharmacological effects such as antidiabetic (4,5), spasmylytic and bronchodilator (6,7), analgesic and anti-inflammatory (8). The extracts also showed in vitro and in vivo antibacterial effects (9). We decided to study the effects of neutral, acidic, basic fraction and essential oil from extracts of the Nigella sativa seed on some pathogenic bacterial strains.

MATERIALS AND METHODS

A. Plant material. Nigella sativa seeds were obtained from local market in Mosul city.

B. Preparation of crude extract. 150 gm of the powdered dried seeds of Nigella sativa were used for the preparation of the aqueous extract (10,11,12).

C. Extraction. We have separated three major fractions namely, neutral (N), acidic (A) and basic (B) from the aqueous extract of Nigella sativa seeds as indicated in Scheme 1.

D. Bacteria. The bacteria species used are listed in Table (1). All strains were obtained from Biology Department, College of Science, University of Mosul. Then growth up to the stationary phase in a nutrient broth at 37 °C and a sample of 0.5 ml of each bacteria was spread over a surface of a Muller-Hinton agar plate (13).

E. Antibacterial assay. Discs of filter paper, 6 mm in diameter, were sterilized at 140 °C for 1 h. and impregnated with 1 ml of stock solution of 10 mg/ml of each fraction and then dried. Distilled water was used as a solvent for the three fractions and dimethyl sulfoxide (DMSO) for the antibiotics and ethylene glycol for essential oil. The inoculated plates were incubated at 37 °C for 18 h and the inhibition zones were measured.
Scheme 1. Extraction procedure for the aqueous extract of *Nigella sativa* seeds
Antibacterial Studies of Essential Oil and Different

F. a. Preparation of Nigella sativa seeds oil. The Nigella sativa seeds oil was prepared according to reported procedure (Khodair et al., 1993) by grinding 50 gm of the seeds in 250 cm³ of ethyl alcohol (95%) in the grinder in side an ice-bath. Shake the mixture by electrical shaker for 1 hr at room temperature filter the suspension through many layers of gauze then through the filtration funnel to get rid from the hard residue. Centrifuge the filtrate with speed of (1000 xg) for 15 minutes. Then collect the oily layer which composed of the oil of the seeds.

b. Steralization of Nigella sativa seeds oil. 1 cm³ of the oil dissolved in 9 cm³ of ethylene glycol then sterilized by using filter membrane (0.22 micron). The sterilized oil used for further dilution used in the experiments.

G. Test of isolation compounds:

a. Alkaloids test (basic fraction). Myers test was used to identified the alkaloids, we mixed 5 ml from sample with drops from the reagent, then we showed white precipitate, that is positive result of alkaloids test (15).

b. Acids test. Iodate-idide test was used to identified the acids, we added 2 drops from 2% potassium iodide to the sample and 2 drops from 4% potassium iodate, after boiling the mixture for 1 minute, added drops from starch solution, then we showed blue colour which referred positive result of acid test (16).

H. Identification of isolation compounds. Identified the isolation compounds by used the infrared (IR) by used the infrared spectrophotometer model Tensor 27 Bruker Co., Germany (17).

RESULTS AND DISCUSSION

The investigation of the acidic and basic extract and the basic oil of the Nigella sativa seeds showed different effects against the bacterial including Staph. aureus, Sal. typhi, P. mirabilis, B. subtilis, Ps. aeruginosa, Sh. Flexneri, Sal. typhimurium which used in the present study as shown in Table (1).

The neutral fraction study did not show any effect against bacteria used in the present study. As well the acidic and basic extracts with basic oil showed no effect against B. subtilis and Ps. aeruginosa (Table 1). The basic oil showed almost similar inhibition effect against both bacteria S. typhimurium, S. typhi and Sh. flexneri compared with the standard antibiotic Gentamycin, and high inhibition effect compared with the Amoxicillin so the diameter of the inhibition zone at the concentration 0.1 cm³/cm³ was 22 mm, while the diameter of the inhibition zone of the antibiotic at concentration 10 mg/cm³ was 21 mm. the acidic extract for the N.S. seeds showed less effect compared with basic oil against the
bacteria mentioned above. So, the diameter of the inhibition zone at concentration 10 mg/cm³ for the bacteria *Sal. typhimurium* was 19 mm, while the effect of acidic extract against the *Sal. typhimurium* were more than that for the standard antibiotics Gentamycin and Amoxicillin (18,19). The effect of the basic extract against those bacteria mentioned above were less than that for both the basic oil and the acidic extract. The examination results showed that the effect of the basic oil and both extracts against the *Staph. aureus*, *E. coli* and *Pr. Mirabilis* were less than that for the other types of bacteria as shown in Table (1). This was further supported by other researchers (20,21,22) in their studies of the basic oil and both, the acidic and basic extracts against the gram positive and gram negative bacteria. We could conclude from this study that the gram negative bacteria were more sensitive toward the basic oil and the both extracts except the *Pseudo. aeruginosa*. Meanwhile, the *Sal. typhimurium* and *Sh. flexnai* were the most sensitive bacteria toward the extracts and this was in accordance with many studies (23).

The extracted compounds were confirmed spectrochemically by using the IR spectroscopy. The absorption of infrared spectroscopy (IR) gave absorption of stretching for (O-H) and (N-H) bands between (3300-3550) cm⁻¹. It showed absorption of (C-H) bonds stretching at 2855 cm⁻¹ and 2927 cm⁻¹, so gave absorption of (C=O) bond stretching at 1724 cm⁻¹, (C=N) bond stretching at 1654 cm⁻¹, (N-H) bond bending at 1557 cm⁻¹, (C-O) bond bending at 1100 cm⁻¹, (C-H) bond bending at 784 cm⁻¹. These all absorptions refer to that these compounds have alkaloidic properties (Figure 1).

The absorption of infrared spectroscopy (IR) showed that these isolated compounds have acidic organic properties which gave absorption at 1446 cm⁻¹ ascribe to (C=O) bond, so gave a broad absorption range (3300-3600) cm⁻¹ ascribe to carboxylic hydroxyl (OH). They showed absorption of (C-O) bond bending between (1004-1250) cm⁻¹ as to aromatic system showed absorptions between (1464-1558) cm⁻¹ acidic (Figure 2).
Table (1): Antibacterial activity of the basic, acid and essential oil of Nigella sativa seeds against tested bacteria. Diameter of inhibition zone (mm)

<table>
<thead>
<tr>
<th>Microorganisms</th>
<th>Concentration (mg/ml)</th>
<th>Concentration (cm³/cm³)</th>
<th>Control 1 mg/ml</th>
<th>Control 2 mg/ml</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Basic fraction</td>
<td>Acidic fraction</td>
<td>Neutral fraction</td>
<td>Essential oil</td>
</tr>
<tr>
<td>Staph. aureus</td>
<td>1 7 1 1 1 1 0 0 0 0 1</td>
<td>1 3 1 8 - - - - - -</td>
<td>14 10 - - - -</td>
<td>1 0 0 0 0 0 1 0</td>
</tr>
<tr>
<td>B. subtilis</td>
<td>1 - - - - - - - - - -</td>
<td>1 7 1 2 0 - - - - -</td>
<td>1 0 1 0 7 1 1</td>
<td>6 0 - - 1 7 - -</td>
</tr>
<tr>
<td>Pr. mirabilis</td>
<td>1 8 2 2 1 1 3 0 0 1 0</td>
<td>1 7 6 1 4 1 0 5 0 9</td>
<td>15 11 1 2 7 1 1</td>
<td>1 0 0 0 0 1 0</td>
</tr>
<tr>
<td>Sal. typhi</td>
<td>1 6 2 8 - - - - - - -</td>
<td>1 7 4 1 0 - - - -</td>
<td>1 1 0 0 0 1 0</td>
<td>1 0 0 0 0 1 0</td>
</tr>
<tr>
<td>E. coli</td>
<td>1 4 1 8 - - - - - - -</td>
<td>1 5 3 1 0 - - - -</td>
<td>1 1 0 0 0 1 0</td>
<td>1 0 0 0 0 1 0</td>
</tr>
<tr>
<td>Ps. aeruginosa</td>
<td>- - - - - - - - - - -</td>
<td>- - - - - - - - - -</td>
<td>- - - - - - - -</td>
<td>- - - - - - - -</td>
</tr>
<tr>
<td>Sal. typhimurium</td>
<td>1 7 3 1 0 - - - - - -</td>
<td>1 9 1 5 12 8 - - - -</td>
<td>23 18 13 7 1 3 1 8 - -</td>
<td>2 1 4 8</td>
</tr>
<tr>
<td>Sh. flexneri</td>
<td>1 5 4 9 - - - - - - -</td>
<td>1 6 2 8 - - - - - -</td>
<td>20 16 10 - - - -</td>
<td>1 4 1 - - - -</td>
</tr>
</tbody>
</table>

(-) Resistant
Figure (1): The IR spectra of the basic extracts
Figure Q: The IR spectra of the acidic extracts
REFERENCES